IMPROVING NUTRITIONAL QUALITY OF THE GOAT MILK BY GRAZING


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Abstract. The aim of study was to investigate the effect of grazing on the nutritional quality, such as composition, vitamins and fatty acids content of dairy goat milk. Before vegetation period, all goats were kept indoors and nutrition based on hay diet. After turn-out to pasture, all goats were grazing. Bulk milk samples were collected on 12 consecutive days at two sampling periods: in indoor and in grazing months and analysed for milk compositions and vitamin A, E and D3, as well as for fatty acid contents. The milk from grazing goats had significantly higher fat, protein and total solids non-fat than goats kept indoors. Grazing caused higher concentrations of vitamin A (0.026 vs. 0.036 mg/100ml; P<0.01) and D3 (0.075 vs. 0.089 mg/100ml; P<0.05) compared to feeding hay. During the grass diet the rumenic acid (0.56 vs. 0.66 g/100g fatty acids; P<0.05) and n-3 fatty acids (0.36 vs. 1.19 g/100g fatty acids; P<0.001) contents in milk significantly increased. In this study, n-6/n-3 ratio of 10.17 and 1.82 were found in milk samples of goats that fed indoor and grass, respectively. It can be concluded that the milk from grazing goats is more advantageous for human nutrition, than the milk produced by animals fed hay based diet.

Keywords: goat, grazing, milk, vitamins, fatty acids

Introduction

Pasture is one of the most natural feeding sources for animals. The digestibility, protein and energy content of the grass are notable, the crude fat content relatively low, 2-3% of dry matter, nevertheless the grass is rich in α-linolenic acid (Cabiddu et al. 2005), accordingly pastures and other green forages diet have been associated with high content of n-3 fatty acids of milk fat. Feeding green forages to dairy animals increased the concentrations of long chain fatty acids, such as α-linolenic acid and reduced linoleic acid and n-6/n-3 ratio in milk and in cheese (Tsiplakou et al., 2006; Pajor et al. 2012). It is well known, that the high ratio of n-6/n-3 fatty acids is a risk factor in coronary heart disease (CHD). The recommend value for the n-6/n-3 ratio is less 4:1 (Simopoulos, 2004).

In addition, the milk possesses other favourable components, such as rumenic acid and fat-soluble vitamins. The importance of goat milk consumption is recently increasing, because the milk compounds are beneficial effect in human diet. Numerous studies on effect of pasture on milk fatty acids, especially on rumenic acid composition were carried out in cattle and sheep (e.g. Frelich et al. 2012 and Tsiplakou et al., 2006). The rumenic acid (c9t11CLA isomer) has a range of positive health properties such as anticarcinogenic (Ip et al., 1991) and antiatherogenic effects (Nicolosi et al., 1997). In contrast, literature reports are limited about goat milk and cheese.
The fat-soluble vitamins are knowingly favourable in human diet for their antioxidant potential and positive impact on health (Bergamo et al., 2003).

Moreover, there is little available information about the content of vitamins in goat milk, especially about pasture kept goats compared to other ruminants (e.g. Kondyli et al. 2007).

Thus, the aim of this study was to investigate the effect of the grazing on certain nutritional quality compounds (vitamin A, E and D₃, n-3 fatty acids and rumenic acid) of goat milk.

Materials and method

Experimental animals and diet

The study was carried out in a goat farm in Borsod-Abauj-Zemplen County (Northeast Hungary). 54 Hungarian Native goats on different parities but in the same stage of lactation were involved in this study. Before grazing months (May), all goats were kept indoors and their nutrition was based on ad libitum alfalfa hay and also 350 g/day grain mix.

After turn-out to pasture the grazing group stayed all day long on the pasture; however, they were also fed with 350 g/day grain mix. Both groups had same composition of the grain mix, which was given twice a day in equal amounts at milking time. A commercial vitamin (A, D₃, E) and trace-mineralized salt block was provided free choice to all goats. The diets were adjusted to the NRC (2007) recommendations of energy and protein requirements for dairy goats (body weight: 60 kg; 2.5 kg of milk/day).

Utilization of native pasture was extensive in order to avoid over-grazing. The stocking density of the pastures grazed by the goats was about 0.5 AU/ha. The main grass and legume species were Festuca pseudovina and Trifolium pratense. Other species were Elymus repens, Elymus hispidus, Bromus inermis, Calamagrostis epigeios and Arrhenatherum elatius. The average annual rainfall of the area is approx. 695 mm. The annual green grass yield was 5.2 t/ha. During the daily routine the grazing goats were driven to pasture after the morning milking, and collected in the afternoon to be milked and confined for the entire night.

Milk samples were taken at two sampling times: in mid-April and in mid-July when the goats were on average among the 30 and 120 days in lactation. Samples of pooled milk were collected twice a day during 12 consecutive days of the experimental months at 6.00 a.m. and 6.00 p.m., all milk samples were frozen and stored at -20 °C until further analysis. Before laboratory analysis, twice a day gathered milk samples were combined to one sample for the analysis of chemical composition.

Chemical analysis

Fat, protein, lactose and total solids without fat contents of milk were determined using a Bentley device (Bentley Combi apparatus, Bentley Instruments Inc, Chaska, MN, USA).

The milk fat was dissolved in sodium hydroxide-methanol solution and re-esterified to methyl-esters according to the AOAC (1990) method using boron trifluoride (BF₃). Methyl esters of fatty acids were determined by gas chromatography using a Shimadzu
GC 2010 apparatus (Japan) with a flame ionization detector (FID) and column (CP-SIL-88, 100 m × 0.25 mm × 0.2 μm). The split injection ratio was 50:1. The column oven temperature was held at 80°C for 0 min, then programmed at a rate of 2.5°C/min up to 205°C and held for 20 min and then increased again to 225°C at 10°C/min, and held for 5 min. The injector and detector temperatures were 270°C and 300°C, respectively. Helium was used as the carrier gas, applying a flow rate 28 cm/s. Peaks were identified on the basis of the retention times of standard methyl esters of individual fatty acids (Mixture Me 100, Larodan Fine Chemicals AB, Sweden). The proportions of the individual acids were calculated by the ratio of their peak area to the total area of all observed acids.

Fat-soluble antioxidants (vitamin A, E and D₃) were analysed using the high-performance liquid chromatographic method described by Kerti and Bardos (2006). The vitamin detection was performed at 325 nm for vitamin A, 290 nm for vitamin E and 265 nm for vitamin D₃. The peak areas were integrated and quantified by using the Chrompass software (Jasco, Japan).

Statistical analysis

Effects of feeding method was analysed as independent variable. Statistical analysis was processed by the SPSS 21.0 software package (Shapiro-Wilk test for normality distribution, F test for equality of Variances, t-test and Welch’s corrected t-test). Significance was taken at an alpha level of 0.05.

Results and discussion

There was no significant difference in the milk production of the examined groups; daily milk yield during the investigation was 2.5 kg in control goats and 2.6 kg in experimental goats.

Milk composition was significantly affected by diet. Hay diet caused lower (P<0.05) fat (3.11 g/100g), protein (2.94 g/100g) and total solids non-fat (7.87 g/100g) composition compared to milk from pasture kept goats (3.70, 3.22 and 8.15 g/100g, no data in Table). Thus, the grazing significantly improved the milk composition during experiment. It is well known, that the goat milk fat and protein content is affected by many factors, such as breed, parity, stage of lactation and diet (Kuchtik et al., 2008; Novotna et al., 2009) and it has great effect on cheese composition. Soryal et al. (2004) in Alpine goat and Pajor et al. (2009) in Hungarian Native goat found that the grazing slightly increased the milk compositions.

The contents of the selected vitamins in the goat milk are presented in Table 1.

<table>
<thead>
<tr>
<th>Items</th>
<th>Control diet</th>
<th>Experimental diet</th>
<th>Total</th>
<th>SEM</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Vitamin A</td>
<td>0.025</td>
<td>0.036</td>
<td>0.031</td>
<td>0.002</td>
<td>0.001</td>
</tr>
<tr>
<td>Vitamin E</td>
<td>0.122</td>
<td>0.140</td>
<td>0.131</td>
<td>0.011</td>
<td>0.454</td>
</tr>
<tr>
<td>Vitamin D₃</td>
<td>0.075</td>
<td>0.089</td>
<td>0.082</td>
<td>0.003</td>
<td>0.017</td>
</tr>
</tbody>
</table>

P: level of significance
The average contents of vitamins A and D$_3$ were 0.031 mg and 0.082 mg/100 ml. In goats reared indoor lower contents of A and D$_3$ vitamins (0.025 mg and 0.075 mg/100 ml of milk) were found compared to goats with experimental diets (0.036 mg and 0.089 mg/100 ml of milk).

Mean of vitamin E concentration was 0.131 mg/100 ml, however vitamin E contents of milk did not show any significant differences between the experimental periods. The mean values of vitamin A and E as well as vitamin D$_3$ in present study were slightly higher compared to values which published by Kondyli et al. (2007) and Raynal-Ljutovac et al. (2008).

Our results showed that milk from grazing goats is a good dietary source of vitamins A, E and D$_3$ whereas these are also well known for their antioxidant potential (Bergamo et al., 2003).

The results of the fatty acid analysis of milk samples are presented in Table 2.

<table>
<thead>
<tr>
<th>Fatty acids</th>
<th>Control diet</th>
<th>Experimental diet</th>
<th>Total</th>
<th>SEM</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>C4:0</td>
<td>0.58</td>
<td>0.71</td>
<td>0.64</td>
<td>0.03</td>
<td>0.037</td>
</tr>
<tr>
<td>C6:0</td>
<td>1.32</td>
<td>1.73</td>
<td>1.52</td>
<td>0.06</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>C8:0</td>
<td>1.68</td>
<td>2.20</td>
<td>1.94</td>
<td>0.08</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>C10:0</td>
<td>7.59</td>
<td>9.28</td>
<td>8.43</td>
<td>0.29</td>
<td>0.001</td>
</tr>
<tr>
<td>C12:0</td>
<td>3.40</td>
<td>3.39</td>
<td>3.40</td>
<td>0.07</td>
<td>0.971</td>
</tr>
<tr>
<td>C14:0</td>
<td>11.27</td>
<td>9.91</td>
<td>10.59</td>
<td>0.21</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>C14:1c9</td>
<td>0.16</td>
<td>0.08</td>
<td>0.12</td>
<td>0.01</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>C16:0</td>
<td>28.76</td>
<td>27.83</td>
<td>28.29</td>
<td>0.26</td>
<td>0.072</td>
</tr>
<tr>
<td>C16:1c9</td>
<td>0.96</td>
<td>0.73</td>
<td>0.84</td>
<td>0.03</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>C18:0</td>
<td>10.71</td>
<td>17.97</td>
<td>14.34</td>
<td>0.81</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>C18:1n-9c</td>
<td>17.89</td>
<td>14.43</td>
<td>16.16</td>
<td>0.58</td>
<td>0.001</td>
</tr>
<tr>
<td>C18:1t11TVA</td>
<td>0.77</td>
<td>2.15</td>
<td>1.46</td>
<td>0.19</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>C18:2n-6</td>
<td>2.89</td>
<td>2.00</td>
<td>2.45</td>
<td>0.10</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>c9t11CLA</td>
<td>0.56</td>
<td>0.66</td>
<td>0.61</td>
<td>0.02</td>
<td>0.021</td>
</tr>
<tr>
<td>C18:3n-3</td>
<td>0.35</td>
<td>1.19</td>
<td>0.77</td>
<td>0.09</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>C20:4n-6</td>
<td>0.24</td>
<td>0.14</td>
<td>0.19</td>
<td>0.01</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>SMCFA</td>
<td>11.17</td>
<td>13.91</td>
<td>12.54</td>
<td>0.44</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>C:12+C:14+C:16</td>
<td>43.43</td>
<td>41.13</td>
<td>42.20</td>
<td>0.49</td>
<td>0.008</td>
</tr>
<tr>
<td>SFA</td>
<td>75.65</td>
<td>78.30</td>
<td>76.98</td>
<td>0.60</td>
<td>0.024</td>
</tr>
<tr>
<td>MUFA</td>
<td>20.25</td>
<td>17.71</td>
<td>18.98</td>
<td>0.55</td>
<td>0.016</td>
</tr>
<tr>
<td>PUFA</td>
<td>4.09</td>
<td>3.99</td>
<td>4.04</td>
<td>0.07</td>
<td>0.477</td>
</tr>
<tr>
<td>n-6</td>
<td>3.18</td>
<td>2.15</td>
<td>2.66</td>
<td>0.12</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>n-3</td>
<td>0.36</td>
<td>1.19</td>
<td>0.77</td>
<td>0.09</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>n-6/n-3</td>
<td>10.17</td>
<td>1.82</td>
<td>6.00</td>
<td>1.04</td>
<td>&lt;0.001</td>
</tr>
</tbody>
</table>

P: level of significance; SMCFA: Short and medium chain fatty acids (C$_4$-C$_{10}$); SFA: saturated fatty acids; MUFA: monounsaturated fatty acids; PUFA: polyunsaturated fatty acids; CLA: conjugated linoleic acid.

Grazing significantly increased the contents of short chain fatty acids [butyric (C4:0), caproic (C6:0), caprylic (C8:0) and capric acids (C10:0)], stearic (C18:0), α-linolenic (C18:3), total saturated fatty acids (SFA) and as well as significantly decreased the
Grazing significantly increased the concentrations of short and medium chain fatty acids (MCFA) in milk. These fatty acids are hydrolyzed rapidly and are absorbed directly to the liver via portal vein (Papamandjaris et al. 1998). Therefore the medium chain fatty acids have been used for patients that have malabsorption syndrome. Recently, the relations between the medium chain fatty acids and certain metabolic syndromes are summarized in a review report by Nagao and Yanagita (2010).

Concentrations of lauric, myristic and palmitic acids were lower in milk samples of the grazing group than in the control group. These acids are known to be hypercholesterolemic, whilst the other major saturated fatty acid (SFA), e.g. stearic acid, does not. Ulbricht and Southgate (1991) reported that these fatty acids to be responsible for increase the level of total and LDL cholesterol concentrations in blood serum.

The grazing positively affected the concentration of rumenic and vaccenic acids in the milk. The rumenic and vaccenic acids concentrations in the milk were 0.56 and 0.77 vs. 0.66 and 2.15% for hay based and experimental diet, respectively. The polyunsaturated fatty acids, such as linoleic and linolenic acid, are partly saturating in the rumen by biohydrogenation. Throughout this process the rumenic acid is formed from linoleic acid in the rumen by anaerobic bacteria (such as B. fibrisolvens), with vaccenic acid (t11C18:1)(TVA) as intermediates. TVA is converted to CLA by Δ⁹-desaturase in mammary gland (Bauman et al. 2001). Because of biohydrogenation in rumen, the relative percentage of C18:0 was significantly increased in milk from grazing goats (17.97%) compared to control group (10.71%).

In present study, grazing significantly decreased the n-6 fatty acids and increased the n-3 fatty acids contents, however the n-6/n-3 ratios were favourable, 1.82 instead of 10.17. The n-6/n-3 ratio is generally used to assess the nutritional value of fats. According to Simopoulos (2004), recommend value for the n-6/n-3 ratio is less 4:1. The low ratio of n-6/n-3 in the milk of grazing goats is meeting with the new recommendations for human nutrition. It is well known, that grass is rich in α-linolenic acid, authors reported that 50-60% of the total fat is n-3 fatty acids in grass (Cabiddu et al. 2005; Tsvetkova and Angelow, 2010). The relatively unfavourable n-6/n-3 ratio in control group was probably due to lower n-3 fatty acids concentrations in hay which is affected by oxidative and leaf losses during hay making (Doreau and Poncet, 2000; Dewhurst et al., 2006). However, the relatively high PUFA content of grass is inhibiting de novo fatty acid synthesis in mammary gland (Couvrer et al., 2006). This confirmed by decrease of miristic and miristoleic fatty acids content of grazed goats’ milk samples.

Conclusions

In conclusion, significant difference was found in milk composition, vitamin as well as fatty acid profile between two treatments. Grazing significantly increased the concentrations of vitamin A and D₃, rumenic acid, short and medium chain fatty acids, as well as n-3 fatty acids in milk. These results show that grazing can enhance the nutrition value and quality of goat milk, consequently, consumers have nutraceutical
benefits from consumption of milk from grazing goats due to higher concentrations of health promoting compounds.

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