

UTILIZATION OF DISTILLERY WASTES FOR SUSTAINING SOIL HEALTH IN PADDY FIELDS

LENINRAJA, D.¹ – RADHA, P.^{2*} – ELANCHEZHIAN, K.³ – RAJINIMALA, N.⁴ – MANOBHARATHI, K.⁵
– SUBASH CHANDRA BOSE, K.⁵

¹*Department of Natural Resource Management, Horticultural College and Research Institute, Tamil Nadu Agricultural University (TNAU), Periyakulam 625604, Tamil Nadu, India*

²*Department of Forest Biology and Tree Improvement, Forest College and Research Institute, Tamil Nadu Agricultural University (TNAU), Mettupalayam 641301, Tamil Nadu, India*

³*Department of Agricultural Entomology, Agricultural College and Research Institute, Tamil Nadu Agricultural University (TNAU), Killikulam, Vallanadu 628252, Tamil Nadu, India*

⁴*Rice Research Station, TNAU, Ambasamudram, Tirunelveli 627401, Tamil Nadu, India*

⁵*Mother Teresa College of Agriculture, Affiliated to TNAU, Illuppur, Pudukkottai 622102, Tamil Nadu, India*

**Corresponding author
e-mail: pradha@tnau.ac.in*

(Received 2nd Aug 2024; accepted 9th Dec 2024)

Abstract. Sugarcane based industries like distilleries produce a large amount of byproducts like Raw Spent Wash (RSW), Treated Distillery Effluent (TDE) and Bio-compost (BC). The Treated Distillery Effluent is waste water, which could be recycled in agriculture both as irrigation water and as a source of plant nutrients. The beneficial effect of organic matter for enhancing the soil fertility and thereby improving the crop productivity is well established. Thus, the soil application of TDE could offer the double benefit of safe disposal of wastes and its effective utilization for agricultural production. Keeping this fact in view, a field experiment was conducted to study the effect of Treated Distillery Effluent (TDE) and bio-compost on soil chemical and biological properties using paddy as a test crop (ADT- 43). Based on the increase in soil chemical and biological properties, it can be concluded that TDE @ 0.1 million L ha⁻¹(M3) or TDE @ 0.15 million L ha⁻¹ (M4) along with 37.5% N as urea + 37.5% N as bio-compost (S7) can be recommended as a nutrient source for paddy crop.

Keywords: *treated distillery effluent, dehydrogenase, phosphatase, urease, bio-compost*

Introduction

Beneficial utilization of industrial effluents, particularly the treated distillery effluent, has been thought of extensively as it contains all the essential nutrients such as nitrogen, phosphorus and potassium besides micronutrients (Manickam, 1996; Umair et al., 2021). Nutrient application in the form of effluent irrigation may be effective in respect of bioavailability of nutrients to plants. Moreover, application of TDE on soils is one of the most economical resources for the soil fertility amelioration due to improvement in soil water holding capacity, texture, structure and nutrients retention. The available nutrient status of the soil is an important property which supports the crop growth while, the crop growth and dry matter accumulation depends upon the ability of the soil to supply nutrients; the nutrients release from the soil in turn depends upon the demands of the growing plants (Valliappan, 1998; Mohamed Haroon and Subash Chandra Bose, 2004; Mishra et al., 2021). Therefore, the main objective of the present study was to

evaluate the effect of different application rates of distillery wastes, on soil chemical and biological properties in paddy field.

Materials and methods

The experimental site lies geographically in between 10°15' and 11°2' North Latitude and 78°10' and 79°5' East Longitude and at an altitude of 90 m above mean sea level. Field experiment was conducted using paddy as a test crop (ADT- 43). The experiment was conducted in a split plot design with four main plots viz., control; TDE @ 0.05 million L ha⁻¹; TDE @ 0.1 million L ha⁻¹; TDE @ 0.15 million L ha⁻¹. Different levels of N fertilizers viz., 100% N as urea, 75% N as urea, 100% N as bio-compost, 75% N as bio-compost, 75% N as urea and 25% N as bio-compost, 37.5% N as urea and 37.5% N as bio-compost and control were imposed as seven subplot treatments and the treatments were replicated twice. TDE was uniformly applied to each plot as per the treatment schedule at 45 days before planting.

The soil of the experimental field belongs to *Typic haplustert*, neutral in pH (pH 7.58) and low in EC (0.30 dSm⁻¹). The organic carbon content (4.00 g kg⁻¹) and the alkaline KMnO₄-N (162 kg ha⁻¹) were found to be low. The Olsen-P level (16 kg ha⁻¹) and the NH₄OAc-K (205 kg ha⁻¹) were medium. The bacterial, fungal and actinomycetes population were found to be 10.2 × 10⁶ CFU g⁻¹ of soil, 14 × 10⁴ CFU g⁻¹ of soil, 5.1 × 10³ CFU g⁻¹ of soil respectively. The urease, phosphatase and dehydrogenase activity were found to be 4.5 µg NH₄-N g⁻¹ dry soil h⁻¹, 10.1 µg p-nitrophenol g⁻¹ of soil h⁻¹ and 2.5 µg TPF g⁻¹ dry soil h⁻¹ respectively. The enzymes viz., Dehydrogenase and urease activities were estimated by Chendrayan et al. (1980), Bremner and Mulvany (1978) and Silva et al. (2013) respectively. The data on various characters studied during the investigation were statistically analyzed by the method given by Gomez and Gomez (1984). The critical difference was worked out at 5% (0.05) probability levels (*Table 1*).

Bio-compost is prepared by utilizing the pressmud (an organic solid material obtained as by-product from sugar industries) as a raw material and the composting process is being carried out by mechanized open window system with the use of treated distillery effluent and bio-inoculants for about 70–80 days (Dinesh et al., 2020). The product is then sun dried, ground and sieved by mechanical separator and finally enriched with bio-fertilizers. The results of bio-compost analysis indicated that it was neutral in reaction (pH 7.56) with considerable amount of salt (EC 6.74 dSm⁻¹). It was rich in organic carbon (21.86%), N (1.58%), P (2.32%), K (4.56%), Ca (2.78%), Mg (1.62%), Na (1.76%) and had traces of micronutrients viz., Zn, Fe, Cu and Mn with a favorable C:N ratio of 20.4 and considerable amount of enzyme activity and microbial population. Hunsigi (2000) and Singh (2008) also reported that bio-compost prepared from distillery effluent have a neutral pH 6.5 to 7.5 (*Table 2*).

The TDE was dark brown in color imparted by melanoidin and had unpleasant odor which may be due to skatole, indole and other sulphur compounds. It was neutral in reaction (pH 7.71), but loaded with high organic and inorganic salts recording high EC (34.6 dS m⁻¹). The total solids, suspended solids and dissolved solids content of the TDE were 51200, 5610 and 45590 mg L⁻¹, respectively. Being originated from plant sources, the TDE was rich in organic carbon (28,500 mg L⁻¹), K (12,650 mg L⁻¹ as K₂O), Ca (2,250 mg L⁻¹), Mg (1,560 mg L⁻¹) with considerable amount of N (2,000 mg L⁻¹), low in P (246 mg L⁻¹), and had relatively small amounts of micronutrients in the order of Fe > Mn > Zn > Cu. The TDE contained large amounts of bases, whose concentration

followed the order of $\text{Ca} > \text{Mg} > \text{Na}$. The BOD and COD of TDE used for land application were 7,890 and 38,562 mg L^{-1} , respectively. The TDE had appreciable counts of fungi, bacteria and actinomycetes. Sodium adsorption ratio, residual sodium carbonate and soluble sodium percentage were below the critical limits, whereas the potential salinity exceeded the critical level as per the irrigation water quality standards (Table 3).

Table 1. Characteristics of initial soil

Parameters	Values
1. Mechanical composition	
Clay (%)	26.75
Silt (%)	6.50
Fine sand (%)	38.88
Coarse sand (%)	25.89
Textural class	Sandy clay
Soil series	Poovalur series
Soil taxonomy	<i>Typic haplustert</i>
2. Physical composition	
Bulk density (Mg m^{-3})	1.35
Particle density (Mg m^{-3})	2.24
Pore space (%)	47.1
Water holding capacity (%)	22.2
3. Chemical composition	
pH (1: 2.5 soil water suspension)	7.58
EC (dSm^{-1}) (1: 2.5 soil water extract)	0.30
CEC [$\text{c.mol.}(p^+) \text{ kg}^{-1}$]	16.40
ESP	9.75
Organic carbon (g kg^{-1})	4.00
Organic matter (g kg^{-1})	6.89
KMnO_4 - N (kg ha^{-1})	162
Olsen - P (kg ha^{-1})	16
NH_4OAc - K (kg ha^{-1})	205
Exchangeable Ca ($\text{cmol } (p^+) \text{ kg}^{-1}$)	9.00
Exchangeable Mg ($\text{cmol } (p^+) \text{ kg}^{-1}$)	5.50
Exchangeable Na ($\text{cmol } (p^+) \text{ kg}^{-1}$)	1.60
Exchangeable K ($\text{cmol } (p^+) \text{ kg}^{-1}$)	0.30
DTPA- Zn (mg kg^{-1})	0.95
DTPA-Cu (mg kg^{-1})	0.68
DTPA-Fe (mg kg^{-1})	11.74
DTPA-Mn (mg kg^{-1})	6.05
4. Biological properties	
Bacteria ($\times 10^6 \text{ CFU g}^{-1}$ of soil)	10.2
Fungi ($\times 10^4 \text{ CFU g}^{-1}$ of soil)	14.0
Actinomycetes ($\times 10^3 \text{ CFU g}^{-1}$ of soil)	5.1
Dehydrogenase activity ($\mu\text{g TPF g}^{-1}$ of soil h^{-1})	2.5
Phosphatase activity ($\mu\text{g p-nitrophenol g}^{-1}$ of soil h^{-1})	10.1
Urease activity ($\mu\text{g NH}_4\text{-N g}^{-1}$ of soil h^{-1})	4.5

Table 2. Characteristics of bio-compost

Sl. No.	Parameters	Values
1	pH	7.56
2	Electrical conductivity (1:2.5) dS m ⁻¹	6.74
3	Organic carbon (%)	21.9
4	Total nitrogen (%)	1.58
5	Total phosphorus (%)	2.32
6	Total potassium (%)	4.56
7	C:N ratio	20.37
8	Total calcium (%)	3.68
9	Total magnesium (%)	2.29
10	Total sodium (%)	1.56
11	Total iron (mg kg ⁻¹)	396
12	Total zinc (mg kg ⁻¹)	87
13	Total copper (mg kg ⁻¹)	41
14	Manganese (mg kg ⁻¹)	410
15	Bacteria (× 10 ⁶ CFU g ⁻¹ of soil)	9.9
16	Fungi (× 10 ⁴ CFU g ⁻¹ of soil)	33.5
17	Actinomycetes (× 10 ² CFU g ⁻¹ of soil)	8.8
18	Dehydrogenase activity (µg of TPF g ⁻¹ of soil h ⁻¹)	10.3
19	Phosphatase activity (µg of PNPP g ⁻¹ of soil h ⁻¹)	18.9
20	Urease activity (µg NH ₄ -N g ⁻¹ of soil h ⁻¹)	8.4

The experimental fields were thoroughly ploughed with tractor drawn iron plough to bring it to a fine tilth. The field was divided into four strips for imposing main plot treatments and each main plot was divided into 14 sub plots of 20 m² to accommodate seven subplot treatments replicated twice. As per the layout, 22 days aged seedlings are transplanted in to the main field from the nursery.

The soil was ploughed well and calculated quantities of Treated Distillery Effluent (TDE) and Bio-compost (BC) were uniformly applied to the concerned plots as per the treatment schedule 45 days before planting and allowed for natural oxidation.

The treatments were allotted randomly using random principle. The buffer channels were provided around each plot. Calculated quantities of TDE and bio-compost were applied to the main and sub plots as per the treatment schedule.

After 30 days, surface of the treated plots were inverted manually with spade to facilitate aeration and oxidation. Then the plots were formed and a fertilizer schedule of 100:50:50 kg N, P₂O₅, K₂O ha⁻¹ was adopted as the 100% recommended dose. Calculated quantities of fertilizers were applied in each treatment as per the treatment schedule. Entire phosphorus dose was applied as basal while nitrogen and potassium were applied in two splits at 25-30, 45-50 days after sowing.

Collection and processing of soil samples

Initial surface (0-25 cm) soil samples were collected prior to the layout of field experiment to assess the initial fertility status of experimental fields. Soil samples were also drawn at 30th (Active tillering stage), 60th (Panicle initiation stage) day and at

harvest stage. The collected soil samples were air dried under shade, powdered with wooden mallet and then sieved through 2 mm sieve and used for chemical analysis. To study microbial population and soil enzymes activity, fresh soil samples were collected separately (Table 4).

Table 3. Characteristics of treated distillery effluent (TDE)

Sl. No.	Parameters	Values
1	Color	Dark brown
2	Specific gravity (g cm ⁻³)	1.18
3	pH	7.68
4	EC (dSm ⁻¹)	36.4
5	BOD (mg L ⁻¹)	7,890
6	COD (mg L ⁻¹)	38,562
7	Total solids (mg L ⁻¹)	51,200
8	Total volatile solids (mg L ⁻¹)	23,320
9	Total suspended solids (mg L ⁻¹)	5,610
10	Total dissolved solids (mg L ⁻¹)	45,590
11	Organic carbon (mg L ⁻¹) (dry weight basis)	28,500
12	Total N (mg L ⁻¹)	2000
13	Total P (mg L ⁻¹)	246
14	Total K (mg L ⁻¹)	12,650
15	Total Ca (mg L ⁻¹)	2250
16	Total Mg (mg L ⁻¹)	1560
17	Total Na (mg L ⁻¹)	730
18	Total Zn (mg L ⁻¹)	15.7
19	Total Fe (mg L ⁻¹)	87.5
20	Total Cu (mg L ⁻¹)	4.95
21	Total Mn (mg L ⁻¹)	10.5
22	Soluble anions (mg L ⁻¹)	
	Cl ⁻	7348.5
	SO ₄ ²⁻	3580.8
	HCO ₃ ⁻	3525.8
23	CO ₃ ⁻	Absent
	Soluble cations (meq.L ⁻¹)	
	K ⁺	226
	Na ⁺	27.2
	Ca ²⁺	48.3
	Mg ²⁺	50.8
24	SAR	3.86
	RSC (meq.L ⁻¹)	-41.3
	SSP	7.71
	PS (meq.L ⁻¹)	245
	Bacteria (× 10 ⁶ CFU ml ⁻¹)	3.5
28	Fungi (× 10 ⁴ CFU ml ⁻¹)	12.2
29	Actinomycetes (× 10 ² CFU ml ⁻¹)	3.5

Table 4. Standard methods followed for the analysis of soil and organic amendments

S.No	Parameter	Method	Reference
1.	Bulk density	Cylinder method	Piper (1966)
2.	Particle density	Cylinder method	Piper (1966)
3.	pH	pH meter, 1:2.5 (soil: water)	Jackson (1973)
4.	EC	Conductivity meter 1:2.5 (soil: water)	Jackson (1973)
5.	Organic carbon	Wet digestion method	Walkley and Black (1934)
6.	Total N	Macrokjeldahl's method	Piper (1966)
7.	Total P	Vanadomolybdate phosphoric yellow color method	Jackson (1973)
8.	Total K	Flame photometric method	Jackson (1973)
9.	Available N	Alkaline permanganate method	Subbiah and Asija (1956)
10.	Available P	NaHCO ₃ extract-colorimetric method	Olsen et al. (1954)
11.	Available K	Flame photometer	Jackson (1973)
12.	CEC	Neutral normal ammonium acetate	Jackson (1973)
13.	Exchangeable Ca	Neutral normal ammonium acetate (Versenate method)	Jackson (1973)
14.	Exchangeable Mg	Neutral normal ammonium acetate (Versenate method)	Jackson (1973)
15.	Exchangeable Na	Flame photometry (neutral normal ammonium acetate)	Jackson (1973)
16.	Exchangeable K	Flame photometry (neutral normal ammonium acetate)	Jackson (1973)
17.	Free CaCO ₃	Volumetric method	Piper (1966)
18.	Available Zn, Cu, Fe and Mn	Atomic absorption spectrophotometry (DTPA extractant)	Lindsay and Norvell (1978)
19.	Microbial population	Serial dilution plate technique	Aneja (2005)

Soil enzyme activity

Dehydrogenase: Fresh composite soil sample (5 g) was taken in a boiling tube along with 1 ml of 3% 2, 3, 5-Triphenyl tetrazolium chloride and 1 ml of 1% glucose. To this 2.5 ml distilled water was added and incubated for 24 h. Then 10 ml methanol was added to the setup and incubated for another 6 h. The sample was then filtered using Whatman No.1 filter paper. The filtrate obtained was red in color. The color developed was read at 485 nm and the concentration of dehydrogenase (X) in the sample was obtained from the standard graph drawn by using Tri Phenyl Formazan (TPF) as standard (Chendrayan et al., 1980).

$$\text{Dehydrogenase activity of the sample } (\mu\text{g of TPF/g}) = X/5$$

Phosphatase: Five gram (w) of the fresh composite soil was taken in a boiling tube with 10 ml distilled water. To this 0.25 ml toluene and 1 ml *p*-nitrophenol phosphate (PNPP) were added and incubated at room temperature for 1 h. Then 5 ml of 0.5 M sodium chloride and 20 ml of 0.5 M sodium hydroxide was added to the sample and filtered through Whatman No.42 filter paper. The color intensity was read at 420 nm and the concentration of phosphatase (X) in the sample was obtained from the standard graph (Tabatabai and Bremner, 1969).

$$\text{Phosphatase activity of the sample } (\mu\text{g of PNPP/g}) = X/w$$

Urease: Ten gram of dry and sieved soil was taken in a 100 ml volumetric flask. To this 1.5 ml of toluene was added, mixed well and incubated for 15 min. Then 10 ml of 10% urea solution and 20 ml of citrate buffer were added, mixed thoroughly, stoppered and incubated for 3 h at 37°C. Then the volume was made up to 100 ml with distilled water, mixed by shaking immediately. The contents were filtered through Whatman No.1 filter paper and 1 ml of filtrate was pipetted out into 50 ml volumetric flask. To this 9 ml of distilled water, 4 ml of phenate and 3 ml of NaOCl were added, mixed well and allowed to stand for 20 min. The volume was made up to 50 ml and mixed well. The bluish green color developed was read at 630 nm. Simultaneously a blank was also run (without urea solution). The concentration of urease in the sample was obtained from the standard graph using diammonium sulphate (Bremner and Mulvany, 1978).

Results

Soil available nitrogen as influenced by different levels of TDE and biocompost

Available nitrogen content was found to be significantly influenced by the TDE and bio-compost application. The available nitrogen content ranged from 142 kg ha⁻¹ in M1S1 at post-harvest stage to 288 kg ha⁻¹ in M3S7 at active tillering stage (Table 5). The available nitrogen content of the soil showed marked decline at post-harvest stage (225 kg ha⁻¹) compared to panicle initiation stage (231 kg ha⁻¹) and active tillering stage (235 kg ha⁻¹).

Among the main plot treatments, application of TDE @ 0.15 million L ha⁻¹ (M4) registered higher available N status of 267 kg ha⁻¹ which was on par with M3 (262 kg ha⁻¹). The control (M1) recorded the lowest value of 168 kg ha⁻¹.

Among the subplot treatments, S6 (application of 75% N as urea + 25% N as compost) recorded higher available N status of 239 kg ha⁻¹ which was being comparable with S7 (37.5%N as urea + 37.5% N as bio-compost) registering 238 kg ha⁻¹. This was followed by 100% N as urea being comparable among themselves with 75% N as urea registering high available nitrogen status of 234 kg ha⁻¹ and 233 kg ha⁻¹ respectively in the soil over rest of the treatments. This was followed by the application of 100% N as bio-compost (227 kg ha⁻¹) which in turn being on par with 75% N as bio-compost (224 kg ha⁻¹) and control recorded the lowest of 215 kg ha⁻¹.

Table 5. Soil available nitrogen (kg ha⁻¹) as influenced by different levels of TDE and bio-compost in paddy

Treatments	Active tillering stage (stage 1)					Panicle initiation stage (stage 2)					Post harvest stage (stage 3)				
	M1	M2	M3	M4	Mean	M1	M2	M3	M4	Mean	M1	M2	M3	M4	Mean
S1	160	221	248	259	222	154	215	244	257	217	142	201	238	249	207
S2	177	232	270	275	238	173	231	268	268	235	170	226	266	260	231
S3	175	234	267	277	238	171	230	265	270	234	168	225	262	264	229
S4	171	227	258	269	231	167	226	256	265	228	163	219	253	256	222
S5	169	225	254	265	228	163	225	252	262	225	162	216	250	253	220
S6	179	240	275	279	243	177	233	273	275	239	175	228	270	268	235
S7	173	229	288	283	243	169	228	282	280	240	165	223	276	273	234
Mean	172	230	265	272	235	168	227	263	268	231	163	219	259	260	225

Treatments	Pooled mean				
	M1	M2	M3	M4	Mean
S1	152	212	243	255	215
S2	173	230	268	267	234
S3	171	230	264	270	233
S4	167	224	255	263	227
S5	165	222	252	260	224
S6	177	234	273	274	239
S7	169	227	282	278	238
Mean	168	225	262	267	230

	Stage	M	S	M at St	S at M	S at St	S at St × M
SEd	3	4	5	6	9	8	16
CD (5%)	6	7	9	12	18	NS	NS

Interaction of main × subplot treatment was found to be significant. Application of TDE @ 0.1 million L ha⁻¹ (M3) along with 37.5% N as urea + 37.5% N as bio-compost (S7) registered high available nitrogen content of 282 kg ha⁻¹ which was found to be comparable with the application of TDE @ 0.15 million L ha⁻¹ (M4) along with 37.5% N as urea + 37.5% N as bio-compost (S7) recording 278 kg ha⁻¹.

Soil available phosphorus as influenced by different levels of TDE and bio-compost

The available P content of the soil showed a progressive decline over stages of crop growth from 19.48 kg ha⁻¹ of tillering stage to 19.05 kg ha⁻¹ at post-harvest stage. Among the main plot treatments, M4 (application of TDE @ 0.15 million L ha⁻¹) registered higher phosphorus availability of 21.36 kg ha⁻¹ which was followed by M3 (application of TDE@ 0.1 million L ha⁻¹) registering 21.19 kg ha⁻¹ while the lower doses of TDE and the control plot recorded low values for M2 (18.52 kg ha⁻¹) and M1 (16.23 kg ha⁻¹) respectively. The above the trend of result was observed at all the stages of the crop growth (Table 6).

Among the sub plot treatments, application of 75% N as urea + 25% N as bio-compost (S6) recorded higher phosphorus availability of 20.19 kg ha⁻¹ which was being comparable with 37.5% N as urea + 37.5% N as bio-compost (S7) which registered 20.05 kg ha⁻¹. This was followed by application of 100% N as urea (S2) which was on par with 75% N as urea (S3) both of which were found superior over the application of 100% N a bio-compost (S4) and 75% N as bio-compost (S5) and control (S1) recorded the lowest (17.97 kg ha⁻¹). The above trend of results was found at all stages of crop growth.

Table 6. Soil available phosphorus (kg ha⁻¹) as influenced by different levels of TDE and bio-compost in paddy

Treatments	Active tillering stage (stage 1)					Panicle initiation stage (stage 2)					Post harvest stage (stage 3)				
	M1	M2	M3	M4	Mean	M1	M2	M3	M4	Mean	M1	M2	M3	M4	Mean
S1	15.35	17.55	20.30	19.90	18.28	15.17	17.16	20.20	20.00	18.13	14.11	16.38	19.90	19.60	17.50
S2	17.20	19.30	21.30	21.40	19.80	16.89	19.11	21.09	22.28	19.84	16.70	18.91	21.00	21.28	19.48
S3	16.70	19.20	21.70	21.20	19.70	16.51	19.01	21.59	22.07	19.80	16.41	18.82	21.40	21.09	19.43
S4	16.15	18.80	20.80	20.60	19.09	15.93	18.43	20.69	21.68	19.19	15.74	18.13	20.50	20.40	18.69
S5	15.95	18.60	20.50	20.40	18.86	15.74	17.94	20.40	21.48	18.89	15.45	17.65	20.30	20.20	18.40
S6	17.45	19.40	22.20	22.30	20.34	17.28	19.21	21.89	22.47	20.21	17.09	19.01	21.79	22.18	20.02
S7	16.50	19.00	22.70	22.85	20.26	16.32	18.72	22.39	22.67	20.02	16.13	18.52	22.29	22.48	19.86
Mean	16.47	18.84	21.36	21.24	19.48	16.27	18.51	21.18	21.81	19.44	15.95	18.21	21.02	21.03	19.05

Treatments	Pooled mean (stages)				
	M1	M2	M3	M4	Mean
S1	14.88	17.03	20.13	19.83	17.97
S2	16.93	19.11	21.13	21.65	19.71
S3	16.54	19.01	21.56	21.45	19.64
S4	15.94	18.46	20.66	20.89	18.99
S5	15.72	18.06	20.40	20.69	18.72
S6	17.27	19.21	21.96	22.32	20.19
S7	16.32	18.75	22.46	22.67	20.05
Mean	16.23	18.52	21.19	21.36	19.32

	Stage	M	S	M at St	S at M	S at St	S at St × M
SEd	0.06	0.07	0.09	0.12	0.18	0.16	0.32
CD (5%)	0.12	0.14	0.18	0.24	0.37	NS	NS

Interaction of main plot × subplot treatment was found to be significant. Application of TDE @ 1.5 lakh L ha⁻¹ (M4) along with 37.5% N as urea + 37.5% N as bio-compost (S7) registered high available phosphorus of 22.67 kg ha⁻¹. This combination was on par with the application of TDE @ 1.0 lakh L ha⁻¹ (M3) along with 37.5% N as urea + 37.5% N as bio-compost (S7) registering 22.46 kg ha⁻¹.

Soil available potassium as influenced by different levels of TDE and bio-compost

Available potassium content of the soil showed a marked decline over the stages of crop growth. The available potassium content ranged from 137 kg ha⁻¹ in M1S1 at post-harvest stage to 428 kg ha⁻¹ in M3S7 at active tillering stage.

Among the main plot treatments, application of TDE @ 0.15 million L ha⁻¹ registered higher potassium availability (404 kg ha⁻¹) which is comparable with the application of TDE @ 0.1 million L ha⁻¹ registering 400 kg ha⁻¹ and the above trend of result was observed at all stages of the growth (Table 7).

Among the subplot treatments, application of 75% N as urea + 25% N as compost, 37.5%N as urea + 37.5% N as bio-compost, 100% N as urea and 75% N as urea being comparable among themselves registering higher available potassium of 337 kg ha⁻¹, 333 kg ha⁻¹, 331 kg ha⁻¹ and 329 kg ha⁻¹ respectively in the soil over the rest of the treatments. Interaction of main plot × subplot treatments was found to be significant. Application of TDE @ 0.15 million L ha⁻¹ (M4) along with 37.5% N as urea + 37.5% N as bio-compost (S7) registered high available potassium (417 kg ha⁻¹). This combination was on par with the application of TDE @ 0.1 million L ha⁻¹ (M3) along with 37.5% N as urea + 37.5% N as bio-compost (S7) registering 416 kg ha⁻¹.

Table 7. Soil available potassium (kg ha⁻¹) as influenced by different levels of TDE and bio-compost in paddy

Treatments	Active tillering stage (stage 1)					Panicle initiation stage (stage 2)					Post harvest stage (stage 3)				
	M1	M2	M3	M4	Mean	M1	M2	M3	M4	Mean	M1	M2	M3	M4	Mean
S1	200	322	392	385	324	182	294	374	370	305	137	263	350	355	276
S2	209	336	409	421	344	198	326	401	415	335	178	296	395	394	316
S3	206	333	413	419	343	194	322	403	412	333	173	293	390	396	313
S4	204	328	408	416	339	190	312	408	408	330	157	279	385	389	303
S5	202	326	403	414	336	187	310	405	405	327	154	273	382	386	299
S6	211	339	424	425	350	200	328	418	418	341	182	302	400	401	321
S7	205	332	427	428	348	192	316	420	420	337	165	288	402	403	314
Mean	205	331	411	415	341	192	315	404	407	330	164	285	386	389	306

Treatments	Pooled mean (stages)				
	M1	M2	M3	M4	Mean
S1	173	293	372	370	302
S2	195	319	402	410	331
S3	191	316	402	409	329
S4	184	306	400	404	324
S5	181	303	397	402	321
S6	198	323	414	415	337
S7	188	312	416	417	333
Mean	187	310	400	404	325

	Stage	M	S	M at St	S at M	S at St	S at St × M
SEd	6	7	9	12	19	16	32
CD (5%)	12	14	19	24	37	NS	NS

Soil phosphatase activity as influenced by different levels of TDE and bio-compost

Application of TDE and bio-compost significantly increased the soil phosphatase activity. Among the main plot treatments, application of TDE @ 0.15 million L ha⁻¹ (M4) recorded highest phosphatase activity of 2.31 µg p-nitrophenol g⁻¹ dry soil h⁻¹ being on par with the application of TDE @ 0.1 million L ha⁻¹ (M3) recording 2.27 µg p-nitrophenol g⁻¹ dry soil h⁻¹. The control (S1) recorded lowest phosphatase activity of 1.40 µg p-nitrophenol g⁻¹ dry soil h⁻¹.

Among the N fertilizer levels, S6 (75% N as urea + 25% N as bio-compost) recorded higher phosphatase activity of 2.06 µg p-nitrophenol g⁻¹ dry soil h⁻¹ which was being comparable with S7 (37.5% N as urea + 37.5% N as bio-compost) recording 2.04 µg p-nitrophenol g⁻¹ dry soil h⁻¹ (Table 8).

Table 8. Soil phosphatase activity as influenced by different levels of TDE and bio-compost

Treatments	Phosphatase activity (µg p-nitrophenol g ⁻¹ soil h ⁻¹)							
	S1	S2	S3	S4	S5	S6	S7	Mean
M1	1.27	1.45	1.43	1.40	1.38	1.49	1.41	1.40
M2	1.81	1.95	1.95	1.90	1.88	1.99	1.93	1.92
M3	2.10	2.31	2.28	2.21	2.17	2.36	2.44	2.27
M4	2.21	2.32	2.34	2.28	2.25	2.38	2.40	2.31
Mean	1.85	2.01	2.00	1.95	1.92	2.06	2.04	1.97

	M	S	M at S	S at M
SEd	0.03	0.01	0.03	0.01
CD (5%)	0.10	0.02	0.10	0.02

The interaction effect of M × S treatment was found to be significant. Application of TDE @ 0.1 million L ha⁻¹ along with 37.5% N as urea + 37.5% N as bio-compost (M3S7) recorded higher phosphatase activity of 2.44 µg p-nitrophenol g⁻¹ dry soil h⁻¹ followed by the application of TDE @ 0.15 million L ha⁻¹ along with 37.5% N as urea + 37.5% N as bio-compost (M4S7) recording higher phosphatase activity of 2.40 µg p-nitrophenol g⁻¹ dry soil h⁻¹.

Soil urease activity as influenced by different levels of TDE and bio-compost

Application of TDE and bio-compost significantly increased the urease activity in soil. Among the main plot treatments, application of TDE @ 0.15 million L ha⁻¹ (M4) recorded highest urease activity of 7.12 µg NH₄-N g⁻¹ dry soil h⁻¹ being comparable with the application of TDE @ 0.1 million L ha⁻¹ (M3) recording 6.98 µg NH₄-N g⁻¹ dry soil h⁻¹. The control recorded the lowest urease activity of 4.33 µg NH₄-N g⁻¹ dry soil h⁻¹ (Table 9).

Table 9. Soil urease activity as influenced by different levels of TDE and bio-compost

Treatments	Urease activity (µg NH ₄ -N g ⁻¹ soil h ⁻¹)							
	S1	S2	S3	S4	S5	S6	S7	Mean
M1	3.91	4.47	4.42	4.31	4.25	4.60	4.36	4.33
M2	5.56	6.02	6.02	5.86	5.80	6.13	5.94	5.90
M3	6.46	7.12	7.03	6.79	6.69	7.25	7.50	6.98
M4	6.81	7.15	7.22	7.03	6.94	7.32	7.40	7.12
Mean	5.68	6.19	6.17	6.00	5.92	6.32	6.30	6.08

	M	S	M at S	S at M
SEd	0.10	0.01	0.10	0.01
CD (5%)	0.30	0.02	0.30	0.02

Among the N fertilizer levels, S6 (75% N as urea + 25% N as bio-compost) recorded high urease activity of 6.32 µg NH₄-N g⁻¹ dry soil h⁻¹ which was being comparable with S7 (37.5% N as urea + 37.5% N as bio-compost) recording 6.30 µg NH₄-N g⁻¹ dry soil h⁻¹. The control (S1) recorded the lowest urease activity of 5.68 µg NH₄-N g⁻¹ dry soil h⁻¹.

The interaction effect of M × S treatment was found to be significant. Application of TDE @ 0.1 million L ha⁻¹ along with 37.5% N as urea + 37.5% N as bio-compost (M3S7) recorded higher urease activity of 7.50 µg NH₄-N g⁻¹ dry soil h⁻¹ followed by the application of TDE @ 0.15 million L ha⁻¹ along with 37.5% N as urea + 37.5% N as bio-compost (M4S7) recording high urease activity of 7.40 µg NH₄-N g⁻¹ dry soil h⁻¹.

Soil dehydrogenase activity as influenced by different levels of TDE and bio-compost

Application of TDE and bio-compost significantly increased the soil dehydrogenase activity. Among the main plot treatments, application of TDE @ 0.15 million L ha⁻¹ (M4) recorded higher dehydrogenase activity of 24.05 µg TPF g⁻¹ dry soil h⁻¹ being comparable with the application of TDE @ 0.1 million L ha⁻¹ (M3) recording 23.54 µg TPF g⁻¹ dry soil h⁻¹. The control recorded the lowest dehydrogenase activity of 14.59 µg TPF g⁻¹ dry soil h⁻¹ (Table 10).

Table 10. Soil dehydrogenase activity as influenced by different levels of TDE and bio-compost

Treatments	Dehydrogenase activity (µg TPF g ⁻¹ soil h ⁻¹)							
	S1	S2	S3	S4	S5	S6	S7	Mean
M1	13.20	15.07	14.90	14.54	14.33	15.39	14.71	14.59
M2	18.74	20.30	20.29	19.80	19.58	20.66	20.03	19.91
M3	21.80	24.01	23.71	22.91	22.57	24.47	25.30	23.54
M4	22.98	24.10	24.35	23.71	23.41	24.68	25.10	24.05
Mean	19.18	20.87	20.81	20.24	19.97	21.30	21.28	20.52

	M	S	M at S	S at M
SEd	0.32	0.01	0.32	0.03
CD (5%)	1.03	0.02	1.03	0.05

Among the N fertilizer levels, S6 (75% N as urea + 25% N as bio-compost) recorded higher dehydrogenase activity of 21.30 $\mu\text{g TPF g}^{-1}$ dry soil h^{-1} which was being comparable with S7 (37.5% N as urea + 37.5% N as bio-compost) recording 21.28 $\mu\text{g TPF g}^{-1}$ dry soil h^{-1} .

The interaction effect of M $\times$ S treatment was found to be significant. Application of TDE @ 0.1 million L ha^{-1} along with 37.5% N as urea + 37.5% N as bio-compost (M3S7) recorded higher dehydrogenase activity of 25.30 $\mu\text{g TPF g}^{-1}$ dry soil h^{-1} followed by the application of TDE @ 0.15 million L ha^{-1} along with 37.5% N as urea + 37.5% N as bio-compost (M4S7) recording higher dehydrogenase activity of 25.10 $\mu\text{g TPF g}^{-1}$ dry soil h^{-1} .

Discussion

Soil available nitrogen

The available N content was significantly influenced by the application of TDE and BC. Among the different treatments, application of TDE @ 0.15 million L ha^{-1} (M4) recorded the highest available N to a tune of 267 kg ha^{-1} which contributed to an increase of 37% over the absolute control. The higher rate of mineralization and release of N from soil, fertilizers and TDE could have contributed for the increase in the available N in the soil.

Among the subplot treatments, S6 (application of 75% N as urea + 25% N as compost) recorded high available N status which contributed to an increase of 10% over check. It was being on par with S7 (37.5%N as urea + 37.5% N as bio-compost) contributing an increase of 9.6% increase over the control. The effect of bio-compost almost at all the stages of observations was found superior to control. This increase could be attributed to release of N upon sustained mineralization of organic manures (Rakkiyappan et al., 2005; Venkatakrishnan and Ravichandran, 2007; Sridharan, 2007; Suganya, 2008; Dinesh, 2011; Selvamurugan et al., 2024).

Among the treatment combinations, application of TDE @ 0.1 million L per hectare (M3) along with 37.5% N as urea + 37.5% N as bio-compost (S7) registered high available nitrogen and was found to be comparable with the application of TDE @ 0.15 million L ha^{-1} (M4) along with 37.5% N as urea + 37.5% N as bio-compost (S7) recording 278 kg ha^{-1} . Higher N availability in soil could be due to the direct contribution of nitrogen supply as well as increased microbial activity due to the added organic matter and partial pressure of carbon dioxide in the effluent treated soil resulting in an enhanced availability of N in soil. Similar results were reported by Satisha (2000), Subash Chandra Bose et al. (2002b), Sridharan (2007) and Stephen et al. (2024). Significant and positive correlation observed between the available N and yield ($r = 0.802^{**}$) also supported the above findings. A marked decline in the available N in the soil was observed with the advancement of crop growth which might be due to the continuous removal of N by the crop, losses due to transformation. However, during the crop growth the reduction in the soil available N could be due to the uptake by growing crop.

Soil available phosphorus

Application of TDE and bio-compost remarkably increased the available P in soil. Among the different treatments, application of TDE @ 0.15 million L ha⁻¹ (M4) had recorded the highest available P to the tune of 21.36 kg P ha⁻¹ which contributed to an increase of 24% over the absolute control. The increase in available P may be due to the application of TDE applied as well as the consequent dissolution of soil mineral P (apatite P). Though TDE was not acidic, its decomposition released organic acids which might have solubilised the soil native P and thus increased the NaHCO₃-P. Similar results were reported by Mallika (2001), Murugaragavan (2002), Maheswari (2011), Dinesh (2011) and Silva and Kardos (2024).

Among the sub plot treatments, application of 75% N as urea + 25% N as bio-compost (S6) recorded higher phosphorus availability of 20.19 kg ha⁻¹ being comparable with 37.5% N as urea + 37.5% N as bio-compost (S7) registering 20.05 kg ha⁻¹, both of which contributed to an increase of 10.9% and 10.4% increase over the check.

Application of TDE @ 0.1 million L ha⁻¹ along with 37.5% N as urea + 37.5% N as bio-compost (M3S7) or 75% N as urea + 25% N as bio-compost (M3S6) being on par with the application of TDE @ 0.15 million L ha⁻¹ along with 37.5% N as urea + 37.5% N as bio-compost (M4S7). The effect of bio-compost almost at all the stages of observations was found superior to check. These results are in line with the findings of Parmer and Sharma (2002) and Singh et al. (2002). The decomposition processes of easily degradable organics might have reduced the binding energy and P sorption capacity of the soil, favoring higher P availability in the soil. Significant and positive correlation observed between the available P and yield ($r = 0.869^{**}$) also supported the above findings. The decline in available phosphorus at harvest stage could be due to crop uptake, physico-chemical transformations of phosphorus (adsorption, precipitation) into insoluble forms or due to microbial immobilization.

Soil available potassium

The available K in the soils was significantly influenced by the application of TDE and bio-compost. Among the main plot treatments, application of TDE @ 0.15 million L ha⁻¹ (M4) registered significantly higher K availability (404 kg ha⁻¹) compared to the other main plot treatments which contributed to an increase of 53.7% over control. The reason attributed to this was due to the high potassium content of TDE (12,650 ppm). Increase in the available K content of the surface soil was sustained even after the harvest. These results also agreed with the findings of Murugaragavan (2002), Janaki (2008), Gbadeyan et al. (2024) and Selvamurugan et al. (2024) who observed that the available K in the soil increased due to the application of effluent.

Among the subplot treatments, application of 75% N as urea + 25% N as compost, 37.5%N as urea + 37.5% N as bio-compost, 100% N as urea and 75% N as urea being comparable among themselves registered higher available potassium content of 337 kg ha⁻¹, 333 kg ha⁻¹, 331 kg ha⁻¹ and 329 kg ha⁻¹ respectively in the soil over the rest of the treatments which contributed an increase of 10.4%, 9.3% and 8.8% increase over control.

The interaction effect of M × S was also significant wherein the application of TDE @ 0.1 million L ha⁻¹ along with 37.5% N as urea + 37.5% N as bio-compost (M3S7) or 75% N as urea + 25% N as bio-compost (M3S6) being on par with the application of

TDE @ 0.15 million L ha⁻¹ along with 37.5% N as urea + 37.5% N as bio-compost (M4S7). The increase may be attributed to the release of mineral K and addition of K rich manures led to the release of K into the soil solution. The present findings fall in line with that of Verma et al. (2002), Prabu (2003), Sivasamy (2004), Venkatakrishnan and Ravichandran (2007) and Dinesh (2011). Significant and positive correlation observed between the available K and yield ($r = 0.748^{**}$) also supported the above findings. The availability of K in the soil got decreased as the crop growth advanced which could be attributed to the uptake of K by the crop.

Soil enzyme activities

Enzyme activity in soil is an indirect indication of the microbial activity, which is directly correlated with soil microbial population. In the present investigation, greater activities of dehydrogenase, urease and phosphatase were associated with the TDE application. The treatment that received TDE @ 0.1 million L ha⁻¹ along with 37.5% N as urea + 37.5% N as bio-compost (M3S7) followed by the application of TDE @ 0.15 million L ha⁻¹ along with 37.5% N as urea + 37.5% N as bio-compost (M4S7) was found to have greater enzyme activities than the control.

The TDE being liquid organic manure increased the organic matter and nutrients content of the soil and subsequently enhanced the microbial biomass. The high dose of TDE along with the recommended dose of NPK recorded the highest value. It implies that organic and inorganic nutrient inputs provided a nutrient rich environment, which is essential for the development of microbes and synthesis of enzymes (Kamalakumari and Singaram, 1995; Tripathi et al., 2022).

Engracia Madejon et al. (2003) found a positive correlation between the organic residues and dehydrogenase, β -glucosidase, urease and protease activities of the soil. Ramana et al. (2002) and Kumar et al. (2019) also reported that the enzyme activities were increased due to the application of distillery effluent. Generally, organic manure addition was found to enhance the microbial activities which in turn favored the synthesis of various enzymes in soil. These three enzymes play a significant role in the bio-transformation of nutrients in soil, and thus influence the nutrients availability in soil and uptake by crops. The mineralization rate of organic P is relevant to both P nutrition of crops and phosphatase activity in soil. Therefore, higher enzyme activities in soil suggested that the mineralization of N and P was greater due to the application of spent wash. Similar result was reported by Dinesh (2020).

Conclusion

In conclusion, this study highlights the potential of TDE being liquid organic manure increased the organic matter and nutrients content of the soil and subsequently enhanced the microbial biomass. The high dose of TDE along with the recommended dose of NPK recorded the highest value. It implies that organic and inorganic nutrient inputs provided a nutrient rich environment, which is essential for the development of microbes and synthesis of enzymes. Thus, it can be concluded that application of TDE @ 0.1 million L ha⁻¹(M3) or TDE @ 0.15 million L ha⁻¹ (M4) along with 37.5% N as urea + 37.5% N as bio-compost (S7) recorded the highest availability of soil available nutrients and enzymatic activity in the soil.

REFERENCES

- [1] Aneja, K. R. (2005): Experiments in Microbiology, Plant Pathology and Biotechnology. – New Age Publishers, New Delhi.
- [2] Bremner, J. M., Mulvany, R. L. (1978): Urease Activity in Soils. – In: Burns, R. G. (ed.) Soil Enzymes. Academic Press, New York, pp. 149-196.
- [3] Chendrayan, K., Adhya, T. K., Sethunathan, N. (1980): Assay of dehydrogenase activity in soils. – Soil Biol. Biochem. 12: 271-273.
- [4] Dinesh, D. (2011): Utilization of distillery industrial wastes as sources of nutrients for maize (*Zea mays* L.). – Ph.D. Thesis, Tamil Nadu Agricultural University, Coimbatore.
- [5] Dinesh, D., Velu, V., Kakade, V., Jinger, D., Murugaragavan, R. (2020): Response of treated distillery effluent application on growth and yield attributes of maize (*Zea mays* L.) under Typic Rhodustalfs of Tamil Nadu. – Annals of Agricultural Research 41(1): pp.29-36.
- [6] Engracia Madejon, S., Burgos, P., Lopez, R., Cabrera, F. (2003): Agricultural use of three organic residues: effect on orange production and on properties of a soil of Spain. – Nutrient Cycl. Agroecosyst. 65: 281-288.
- [7] Gbadeyan, O. J., Sibiya, L., Liganiso, L. Z., Deenadayalu, N. (2024): Waste to energy: the recycling and reuse of sugar industry waste for different value-added products such as bioenergy in selected countries—a critical review. – Biofuels, Bioproducts and Biorefining. <https://doi.org/10.1002/bbb.2579>.
- [8] Gomez, K. A., Gomez, A. A. (1984): Statistical Procedures for Agricultural Research. – John Wiley and Sons, New Delhi.
- [9] Hunsigi, G. (2000): Sugarcane in Agriculture and Industry. – Tata Mc Graw-Hill, New Delhi.
- [10] Jackson, M. L. (1973): Soil Chemical Analysis. – Prentice Hall of India. Pvt. Ltd., New Delhi.
- [11] Janaki. D. (2008): Utilization of distillery spentwash as a manure to crops and its impact on soil, crop and ground water quality. – Ph.D. Thesis, Tamil Nadu Agricultural University, Coimbatore.
- [12] Kamalakumari, K., Singaram, P. (1995): Relationship among soil chemical, biochemical properties and enzyme activities. – Madras Agric. J. 82: 69-70.
- [13] Kumar, V., Sharma, D. C. (2019): Distillery Effluent: Pollution Profile, Eco-Friendly Treatment Strategies, Challenges and Future Prospects. – In: Arora, P. (ed.) Microbial Metabolism of Xenobiotic Compounds. Springer, Singapore, pp.337-357.
- [14] Lindsay, W. L., Norvell, W. A. (1978): Development of DTPA soil test for Zinc, iron, manganese and copper. – Soil Sci. Soc. Am. J. 42: 421-428.
- [15] Maheswari, K. (2011): Eco-friendly and effective nitrogen management with treated distillery effluent and its impact on rice yield and soil properties. – M.Sc., Thesis, Tamil Nadu Agricultural University, Coimbatore.
- [16] Mallika, S. (2001): Ecofriendly utilization of distillery spentwash for enhancing soil fertility and crop productivity. – M.Sc Thesis., Tamil Nadu Agricultural University, Coimbatore, India.
- [17] Manickam, T. S. (1996): – In: Proceedings of National Symposium on Use of Distillery and Sugar Industry Wastes in Agriculture. Oct. 28th & 29th held at ADAC & RI, Tiruchirappalli, Tamil Nadu Agricultural University, pp 14-22.
- [18] Mishra, D., Kumari, S., Jaiswal, A., Arya, S., Yadav, B. R., Thul, S. T., Kumar, S., Pandey, R., Kumar, R., Pandey, A. (2021): Evaluation of distillery sludge as a soil amendment for improving soil quality and sugarcane (CO-265) yield. – Environmental Technology & Innovation 23: 101624.
- [19] Mohamed Haroon, A., Subash Chandra Bose, M. (2004): Use of distillery spentwash for alkali soil reclamation, treated distillery effluent for fertigation of crops. – Indian Farm. 53: 48-51.

- [20] Murugaragavan, R. (2002): Distillery spent wash on crop production in dryland soils. – M.Sc. Thesis, Tamil Nadu Agricultural University, Coimbatore.
- [21] Olsen, S. R., Cole, C. V., Watanabe, F. S., Dean, A. L. (1954): Estimation of available phosphorous in soils by extraction with sodium bicarbonate. – USDA Circular No. 939.
- [22] Parmer, D. K., Sharma, V. (2002): Studies on long-term application of fertilizers and manure on yield of maize-wheat rotation and soil properties under rain-fed conditions in Western Himalayas. – J. Indian Soc. Soil Sci. 50: 311-312.
- [23] Piper, C. S. (1966): Soil and Plant Analysis. – Inter Science Publication Inc., New York.
- [24] Prabu, P. C. (2003): Bioremediation of paper mill effluent polluted habitat. – Ph.D. Thesis, Tamil Nadu Agricultural University, Coimbatore.
- [25] Rakkiyappan, P., Gopalasundaram, P., Radhamani, R. (2005): Recycling of sugar and distillery industry wastes by composting technology. – In: 37th Meeting of Sugarcane Research and Development Workers of Tamil Nadu, pp 24-26.
- [26] Ramana, S., Biswas, A. K., Kundu, S., Sana, J. K., Yadava, R. B. R. (2002): Effect of distillery effluent on seed germination in some vegetable crops. – Bioresource Technol. 82: 273-275.
- [27] Satisha, G. C. (2000): Bioconversion of sugar industrial effluent wastes into enriched compost and its effect on soils and crops. – Ph. D. Thesis. Tamil Nadu Agricultural University, Coimbatore.
- [28] Selvamurugan, M., Doraisamy, P., Maheswari, M., Valliappan, K. (2024): An integrated approach to achieve a circular bioeconomy in the treatment and disposal of distillery spentwash. – Biomass Conversion and Biorefinery. <https://doi.org/10.1007/s13399-024-05581-7>.
- [29] Silva, L. M. D. C., Salgado, A. M., Coelho, M. A. Z. (2013): Urease Activity. – In: Vermelho, A. B., Couri, S. (eds.) Methods to Determine Enzymatic Activity. Bentham Science, Potomac, MD, pp. 292-319.
- [30] Silva, L. R. B., Kardos, L. (2024): Composting of distillery spent wash. – Journal of Environmental Geography 17(1-4): 15-28.
- [31] Singh, N. (2008): Biocompost from sugar distillery effluent: effect on metribuzin degradation, sorption and mobility. – Pest Management Science: formerly Pesticide Science 64(10): 1057-1062.
- [32] Singh, S., Singh, R. N., Prasad, J., Kumar, B. (2002): Effect of green manuring, FYM, and bio-fertilizer in relation to fertilizer nitrogen on yield and major nutrient uptake by upland rice. – J. Indian. Soc. Soil Sci. 50: 313-314.
- [33] Sivasamy, N. (2004): Phytoremediation potential of fodder crops in paper mill effluent polluted soil habitat. – M.Sc., Thesis, Tamil Nadu Agricultural University, Coimbatore.
- [34] Sridharan. B. (2007): Recycling of post methanated distillery spentwash in the soils of Vasudevanallur for maize crop. – M.Sc. Thesis, Tamil Nadu Agricultural University, Coimbatore.
- [35] Stephen, G. S., Shitindi, M. J., Bura, M. D., Kahangwa, C. A., Nassary, E. K. (2024): Harnessing the potential of sugarcane-based liquid byproducts-molasses and spentwash (vinasse) for enhanced soil health and environmental quality. A systematic review. – Frontiers in Agronomy 6: 1358076.
- [36] Subash Chandra Bose, M., Baskar, M., Kayalvizhi, C., Gopal, H., Sivanandham, M. (2002b): Utilization of Distillery Industrial Wastewater as a Source of Plant Nutrients for Rice Production in the East Coast of Tamil Nadu. – In: Extended Summaries, NCCAR, pp 91-92.
- [37] Subbiah, B. V., Asija, C. L. (1956): A rapid procedure for estimation of available nitrogen in soils. – Curr. Sci. 25: 259-260.
- [38] Suganya, K. (2008): Recycling options of one time application of distillery spentwash to maize and groundnut and its impact on soil fertility and groundwater quality. – Ph.D. Thesis, Tamil Nadu Agricultural University, Coimbatore.

- [39] Tabatabai, M. A., Bremner, M. J. (1969): Use of p-nitrophenol phosphate for assay of soil phosphatase activity. – *Soil Biol. Biochem.* 1: 301-307.
- [40] Tripathi, S., Purchase, D., Al-Rashed, S., Chandra, R. (2022): Microbial community dynamics and their relationships with organic and metal pollutants of sugarcane molasses-based distillery wastewater sludge. – *Environmental Pollution* 292: 118267.
- [41] Umair Hassan, M., Aamer, M., Umer Chattha, M., Haiying, T., Khan, I., Seleiman, M. F., Rasheed, A., Nawaz, M., Rehman, A., Talha Aslam, M., Afzal, A. (2021): Sugarcane distillery spent wash (DSW) as a bio-nutrient supplement: a win-win option for sustainable crop production. – *Agronomy* 11(1): 183.
- [42] Valliappan, K. (1998): Recycling of distillery spent wash—an eco-friendly effective reclamation technology for sodic soils. – Ph.D. Thesis., Tamil Nadu Agricultural University, Coimbatore, India.
- [43] Venkatakrishnan, D., Ravichandran, M. (2007): Effect of integrated nutrient management on growth and yield of sugarcane. – *International J. Tropical. Agric.* 25: 1.
- [44] Verma, T. S., Suri, V. K., Paul, J. (2002): Prescription-based fertilizer recommendations for rice, maize and wheat in different agro-climatic zones of Himachal Pradesh. – *J. Indian. Soc. Soil Sci.* 50: 272-277.
- [45] Walkley, A., Black, C. A. (1934): An estimation of methods for determining organic carbon and nitrogen in soils. – *J. Agric. Sci.* 25: 598-609.